**Bioactive Glass and Vitoss Bone Graft Substitute**

Matthew Swift MS\(^1\) and Annie Reza PhD\(^2\)

1-Stryker Trauma and Extremities, Mahwah, NJ
2-Stryker Orthobiologics, Malvern, PA

**Introduction**

Larry Hench, of the University of Florida, is credited with discovering bioactive glass in the late 1960s. Bioactive glass is a surface reactive glass-ceramic biomaterial originated from a research proposal funded by the US Army Medical Research and Development Command. The hypothesis of this initial research was\(^1\):

“The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body”

This research resulted in a 1971 article describing the in vivo interaction of bioactive glass with bone\(^2\). Since this initial publication, four decades worth of research have produced over 1000 articles on the topic of bioactive glass. Many of these articles describe the mechanisms by which bioactive glass can stimulate bone formation. Recent research on the specific formulation of bioactive glass used in the bioactive forms of the Vitoss Bone Graft Substitute has demonstrated its ability to stimulate cellular activity in vitro and improve healing in vivo\(^3,4\).

**Combeite 45S5 Bioactive Glass**

Stryker’s proprietary bioactive glass, Combeite 45S5, is comprised of 45 mol% SiO\(_2\), 25 mol% CaO, 25 mol% Na\(_2\)O and 5 mol% P\(_2\)O\(_5\)\(^5\). Upon implantation, 45S5 bioactive glass releases soluble ionic species include Si\(^+\), Ca\(^{2+}\), and Na\(^+\) into the surrounding environment\(^6\).

This ionic dissolution initiates a series of reactions leading to the deposition of a hydroxycarbonate apatite layer on the surface of the bioactive glass which facilitates bone forming osteoblast cell attachment to the surface\(^7\). Additionally, these ions have been shown to elicit a cellular response including increased cell proliferation and enhancement of the osteogenic phenotype\(^8\).

**In Vitro Testing of Combeite 45S5**

Testing was performed to investigate the in vitro response of human mesenchymal stem cells (hMSC) and primary osteoblasts to the presence of dissolution products of Combeite 45S5 bioactive glass (particle size mix of 32-90 µm and 90-150 µm). Cells exposed to dissolution products were compared to cells cultured in a control medium without bioactive glass dissolution products. When compared to cells cultured under control conditions, hMSC and primary osteoblasts cultured in glass-eluted medium demonstrated increased cellular proliferation (Figure 1), increased collagen elaboration, and increased mineralization (Figure 2)\(^9\). The increased mineralization and collagen elaboration seen in the hMSC group is particularly notable as it indicates a potential for the bioactive glass dissolution products to influence cells towards osteogenic differentiation in the absence of traditional differentiation reagents or exogenous growth factors.

Additionally, culture in the presence of bioactive glass dissolution products further enhanced the native osteogenic phenotype of primary osteoblasts. This was demonstrated by greater deposition of collagen and mineralized matrix relative to untreated controls.

**Figure 1**

Proliferation of hMSC (A) and osteoblasts (B) significantly increased in response to culture in bioactive glass-dissolution media. (n=4, p<0.05)\(^9\)

**Figure 2**

Alizarin Red mineralization staining of calcium deposition by hMSC (Day 28) and osteoblasts (Day 14) demonstrates increased mineralization in the presence of glass dissolution medium compared to glass-free controls.\(^9\)

---

\(^{1}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{2}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{3}\) hench, l. (1971). the human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. bone contains a hydrated calcium phosphate component, hydroxyapatite [ha] and therefore if a material is able to form a ha layer in vivo it may not be rejected by the body.

\(^{4}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{5}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{6}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{7}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{8}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.

\(^{9}\) Hench, L. (1971). The human body rejects metallic and synthetic polymeric materials by forming scar tissue because living tissues are not composed of such materials. Bone contains a hydrated calcium phosphate component, hydroxyapatite [HA] and therefore if a material is able to form a HA layer in vivo it may not be rejected by the body.
In Vivo Testing of Combeite 45S5

Bioactive forms of Vitoss combine Combeite 45S5 bioactive glass (90-150 μm particle size) with a composite of highly porous beta-tricalcium phosphate (β-TCP) and bovine Type I collagen. A canine model using a critical sized defect in the proximo-lateral humeri was used to compare a bioactive form of Vitoss to a form consisting only of the β-TCP and collagen. While both groups demonstrated successful bony in-growth and resorption of the implant over the 52-week study period, mechanical testing of the defect site revealed a 6-12 week faster return to native strength for the bioactive group when compared to the control (Figure 3A). The Vitoss Bioactive group was 30% stronger than the control at 12 weeks (although not statistically significant) and 23% stronger at 24 weeks (statistically significant; p<0.05). Histological analyses showed that the faster restoration of strength may have been the result of increased interconnectivity of new bone growing within the Vitoss Bioactive group, demonstrated by bridging of new bone between β-TCP scaffold morsels (Figure 3 B, C), ultimately resulting in a stronger trabecular network throughout the defect site.

Conclusion

The ability of bioactive glass to impact bone regeneration has been extensively studied over the last four decades, and well-established data demonstrates the chemical properties of glass facilitate a cellular response. Research on Stryker's proprietary formulation of bioactive glass used in bioactive forms of Vitoss Bone Graft Substitute supports the literature and demonstrates and compared to glass-free controls, the presence of bioactive glass:

- Increases cellular proliferation in vitro.
- Enhances the osteogenic phenotype in vitro.
- Increases the rate of bone growth in a animal model.

Further research is needed to confirm how these findings may translate to clinical applications of Vitoss containing bioactive glass.

References


The bioactive effect of Vitoss Bioactive products have not been studied in any human clinical evaluation and the results from in vitro studies may not be predictive of human experience. Vitoss Bioactive Bone Graft Substitutes are intended for use as a bone void filler for voids or gaps that are not intrinsic to the stability of the bony structure. Vitoss Bioactive Bone Graft Substitutes are indicated for use in the treatment of surgically created osseous defects or osseous defects created from traumatic injury to the bone. Vitoss Bioactive Bone Graft Substitutes are intended to be used for filling bony voids or gaps of the skeletal system (i.e., the extremities, pelvis, and posterolateral spine) and may be combined with autogenous blood, and/or bone marrow. Following placement in the bony void or gap, the scaffold resorbs and is replaced with bone during the healing process. The use of Vitoss Bioactive Bone Graft Substitutes are contraindicated in the presence of one or more of the following clinical situations: growth plate fractures, segmental defects, conditions where the surgical site may be subjected to excessive impact or stresses, including those beyond the load strength of fixation hardware, significant vascular impairment proximal to the graft site, metabolic or systemic bone disorders that affect bone or wound healing, infected sites, osteomyelitis at the operative site, defect site stabilization is not possible, intraoperative soft tissue coverage is not planned or possible, in direct contact with the articular space, conditions in which general bone grafting is not advisable. Vitoss Bioactive Bone Graft Substitutes must not be used in patients with a history of anaphylaxis, history of multiple allergies, known allergies to bovine collagen, or who are being treated for desensitization to meat products because this product contains bovine collagen. A surgeon must always rely on his or her own professional clinical judgment when deciding whether to use a particular product when treating a particular patient. Stryker does not dispense medical advice and recommends that surgeons be trained in the use of any particular product before using it in surgery. The information presented is intended to demonstrate the breadth of Stryker product offerings. A surgeon must always refer to the package insert, product label and/or instructions for use before using any Stryker product. Products may not be available in all markets because product availability is subject to the regulatory and/or medical practices in individual markets. Please contact your Stryker representative if you have questions about the availability of Stryker products in your area.

Stryker Corporation or its divisions or other corporate affiliated entities own, use or have applied for the following trademarks or services marks: Stryker. All other trademarks are trademarks of their respective owners or holders. AlloWrap is a registered trademark of AlloSource.